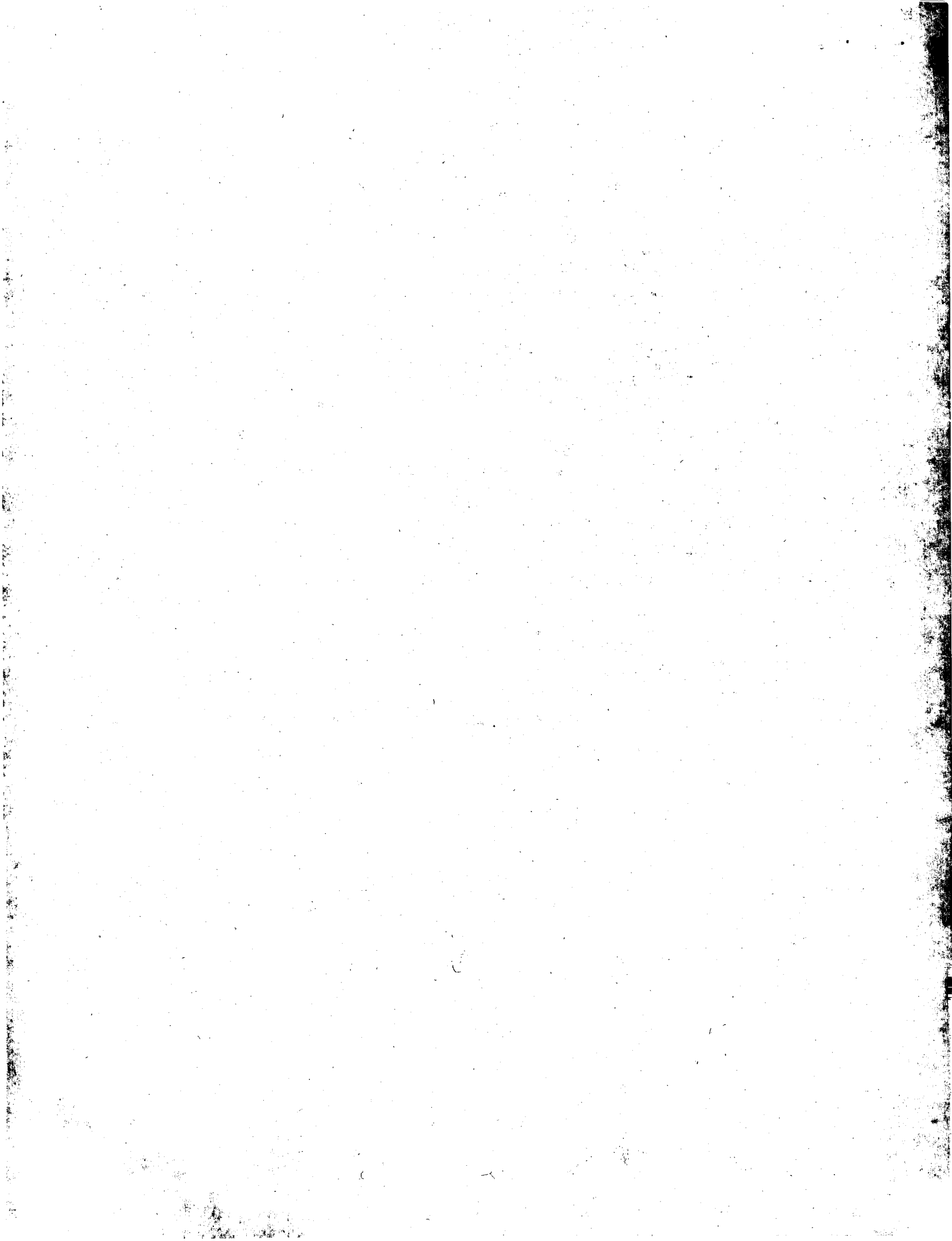


Family list**11** family members for:**EP0317001**

Derived from 9 applications.

- 1 AUF ULTRAKLEINE KOLLOIDALE METALLTEILCHEN
GEGRUENDETES, ALLGEMEIN ANWENDBARES NACHWEISSYSTEM.**
Publication info: **AT85433T T** - 1993-02-15
- 2 Universally Applicable Detection System Based on Ultra Small
Colloidal Metal Particles**
Publication info: **CA1335254 C** - 1995-04-18
- 3 A new universally applicable detection system based On ultra small
colloidal metal particles.**
Publication info: **DE3878167D D1** - 1993-03-18
- 4 A new universally applicable detection system based On ultra small
colloidal metal particles.**
Publication info: **DE3878167T T2** - 1993-05-27
- 5 A new universally applicable detection system based On ultra small
colloidal metal particles.**
Publication info: **EP0317001 A1** - 1989-05-24
EP0317001 B1 - 1993-02-03
- 6 A new universally applicable detection system based On ultra small
colloidal metal particles.**
Publication info: **ES2039025T T3** - 1993-08-16
- 7 A new universally applicable detection system based On ultra small
colloidal metal particles.**
Publication info: **GR3007112T T3** - 1993-07-30
- 8 DETECTION SYSTEM BASED ON SUPERFINE COLLOIDAL METAL
PARTICLE**
Publication info: **JP1201160 A** - 1989-08-14
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- 9 Universally applicable detection system based on ultra small colloidal
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Description

During the last years, small size metal particles, in particular colloidal gold particles, have increasingly been used for the detection and/or quantitative determination of specific binding agents and/or their corresponding bindable substances. Usually, such bindable substances are of biological origin and their determination by means of metal labelling has proven useful in various areas of biochemistry, pharmacology, cytology and histology. The concerned techniques have found widespread use and are particularly attractive for diagnostic purposes.

In principle, the use of colloidal metal particles as markers is based on the fact that the specific binding agent or any substance bindable thereby, when brought into contact with the colloidal metal particles under appropriate conditions, do strongly absorb thereon without losing their affinity for their binding or bindable counterpart.

Mostly, the relationship between a specific binding agent and its corresponding bindable substance(s) will be of the type, antigen-antibody, antibody-antigen, protein-protein, protein-ligand, receptor-ligand or nucleic acid-complementary nucleic acid. Hence, the thus labelled specific binding agents or bindable substance, when allowed to interact with their counterparts will attach their label to the complex formed during the interaction and consequently the detection thereof can easily be performed making use of the properties of the colloidal metal particles.

Depending on the circumstances, colloidal metal particles can be detected, e.g., by direct visual examination, by microscopic or spectrophotometric techniques. A description of "the immunogold staining (IGS) technique", "the sol particle immuno assay (SPIA) technique" of specific applications and improvements thereof can be found in, e.g., U.S. Pat. Nos. 4,313,734, 4,446,238, 4,420,558, 4,775,636, and 4,752,567, and in IBRO handbook series, Wiley, New York, 1983, pages 347 to 372.

So far colloidal metal markers with a wide variation in particle sizes have been prepared. Typical examples are described in Techniques in Immunochemistry Vol. 2 (1983) and in the Journal of Histochemistry and Cytochemistry 35, 1191-1198 (1987) and the references described therein. Usually they are applied in a size range between 5 and 20 nm, however some small size colloidal metal markers of about 2.6 nm have been described in Histochemistry (1985) 83: 409-411. Detection systems based on small colloidal metal markers have a number of advantages over specific binding agents marked with larger metal particles. More particularly they give a higher labelling density and offer an increase in resolution for Elec-

tron Microscopy. However, detection systems based on these small colloidal metal particles have hardly been studied due to their instability and lack of reproducibility. It is therefore an object of the invention to provide reproducible detection systems based on stable and bio-compatible ultra small metal particles.

The present invention provides an aggregate for determining qualitatively or quantitatively bindable substances comprising a colloidal metal particle and a specific binding agent for said bindable substance characterized in that the mean diameter of the colloidal metal particles is ranging between 2 and 0.8 nm. In a further aspect the present invention provides an aggregate for determining bindable substances comprising a colloidal metal particle and a specific binding agent for said bindable substance characterized in that the size of the metal particle is selected so that said aggregate is capable of penetrating into standard biological specimens.

In still a further aspect of the invention there is provided a method for determining a bindable substance using the above identified aggregate.

The aggregate of the present invention is particularly suited for the determination of immunochemical components, such as, haptens, antigens and antibodies.

The colloidal particles of the present invention are either colloidal gold, silver, platinum, palladium, thallium, copper or nickel particles. Preferred examples of particles include gold, silver and platinum particles, with gold particles being the most preferred. Further, the particles may be radioactive, that is, radioactive gold particles, radioactive silver particles and radioactive platinum particles and the like.

In general the preparation of ultra small colloidal metal particles of the present invention is based on the homogeneous reduction of a suitable amount of an appropriate salt, acid or complex of the required metal with a strong reductant at low temperature so as to cause the salt, acid or complex to be reduced to form ultra small colloidal metal particles which are dispersed in a dispersion medium, such as water, straight chain or branched alcohols each having 1 to 10 carbon atoms, e.g. ethanol or methanol, water-miscible ethers e.g. dioxane or diethyl ether, and mixtures thereof. Preferred metal salts, acids or complexes include potassium tetrachloro-platinate, silver nitrate, chloroauric acid and the like salts, acids and complexes, or their radioactive isotopes. An effective reducing agent in the process is phosphorous, in particular white phosphorous, which is added in a stoichiometric amount or a slight excess. Suitable conditions for this process include a pH value between about 2 and 11, a temperature between 0 and 10°C, prefer-

ably below 4°C and vigorous stirring, varying between 100-1000 r.p.m. The reduction reaction may be instantaneous or may last as long as 10 minutes and is indicated by the appearance of a specific colour, such as, for example, brown in case of colloidal gold.

In a presently preferred embodiment the homogeneous reduction is effected at a temperature between 0-4°C by i) preparing an aqueous solution of an appropriate salt, acid or complex of the required metal; ii) preparing an aqueous suspension of phosphorous by mixing water with an appropriate amount of a saturated solution of phosphorous in a lower alkanol, e.g. ethanol or an ether, e.g. diethyl ether; iii) quickly adding one of the solutions to the other under vigorous stirring to form a colloidal sol; iv) and, if desired, removing the excess of reductant and/or the lower alkanol or ether from the sol at a low temperature, preferably at a temperature below 10°C, optionally under reduced pressure.

In particular, ultra small colloidal gold particles may be prepared by mixing during a limited period of time at a temperature between 0 and 4°C, one volume unit of 0.005-0.01% tetrachloroauric acid solution and one volume unit of an aqueous suspension of white phosphorous. The latter being prepared by addition of 0.005-0.0075 volume units of a clear and saturated solution of white phosphorous (4°C) in ethanol to approximately one volume unit of distilled water. Subsequently the excess amount of reductant is removed at a temperature below 10°C under reduced pressure after addition of a sufficient amount of base to convert the remaining phosphorous to phosphine, if desired, the remaining traces of reductant can be removed by oxidation by passing air through the sol, or by addition of an oxidant, e.g. hydrogen peroxide.

Measurement of the colloids by use of a transmission type of electron microscope of model Philips EM 420 shows that the mean diameter of the ultra small colloidal metal particles in the thus prepared colloidal metal sol is ranging between 0.8 and 2 nm, in particular between 1 and 1.8 nm, preferably below 1.6 nm and that the size distribution is bell curved. The variation coefficient being less than 20%, usually less than 15%, in particular between 12% and 6%.

It is contemplated that the thus prepared colloidal metal particles have a crystalline core and consist of at least three metal atoms. It is further assumed that the preferred colloidal metal particles contain between 12 and 600 metal atoms.

The attachment to specific agents or any agents bindable thereby and the various methods of combining them, directly or indirectly, with the desired bindable substances is similar to those

methods known in the art. In this connection reference is made to U.S. Pat. Nos. 4,313,734 and 4,775,636; immunohisto-chemistry, Cuellar, A.C. (ed.), IBRO handbook series, Wiley, New York, 1983, pages 347 to 372, Techniques in Immunocytochemistry Vol. 2, pages 217 to 284 (1983) and Immunocytochemistry, Wright, Bristol, 1986 pages 115 to 146.

For example, the attachment of the specific binding agent is easily effected by contacting the ultra small metal particles preferably at a temperature between 0 and 10°C with an aqueous medium of appropriate pH wherein the desired binding agents, e.g., antibodies, are dissolved. An appropriate pH being between 4.5 and 10. In order to protect the particles from non-selective interactions with non-specific proteins of the test samples it may be appropriate to add quenching or stabilizing agents such as, for example, immunochemically inert polar macromolecules, e.g., bovine serum albumin (BSA), polyvinyl pyrrolidone (PVP) and polyethylene glycol (PEG). After a suitable period of time the aggregates can be concentrated and dialysed via art-known procedures e.g. vacuum dialysis.

The thus prepared aggregates are relatively stable and reproducible when compared with the unbound ultra small colloidal particles especially when the desired binding agents e.g. antibodies and/or their fragments are attached to the colloids immediately after the preparation of the sol. This property can easily be demonstrated by measurements of the particle size with electron microscopy or by the absence of changes in the absorption spectrum in the visible spectrum ranging between 480 and 540 nm.

It was surprisingly found that the mean ratio between the colloidal metal particles and the binding agents in the thus prepared aggregates is at least one, and in particular about 2 which property was indicated by the statistical analysis of the single vs multiple particles frequency of aggregates in electron micrographs.

Another point which should be emphasized in comparison with the prior-art detection systems is the ability of the present aggregates to penetrate into standard biological specimens, in particular into cryosections and chemically fixed intact cells. Probes based on 3nm particles and larger do not penetrate into standard biological specimens, unless after extensive treatment of the preparates with detergents, in which the biological structural coherence is strongly destroyed.

An important factor which co-determines the penetrative capacities of an aggregate is the physical diameter of the aggregate formed by the metal particle and the binding agent. The hydrodynamic radius of the colloidal metal particle is about 50% larger than the actual physical diameter of the solid

particle. It is known that the hydrodynamic radius is decreased after absorption of the specific binding agent, which implies that for aggregates with gold particles smaller than 5 nm the actual radius is largely determined by the radius of the binding agent itself. This in turn means that the penetrative capacity of a "3-5 nm aggregate" and a "1 nm aggregate" should not differ essentially.

In contrast herewith, there has now been found that immunoglobulin (IgG) coupled with one or more ultra small colloidal particle(s) does penetrate into cryosections whereas 4 nm gold particles coated with protein A do not.

The aggregates of the present invention do not only penetrate into cryosections but also penetrate into chemically fixed intact cells being optionally permeabilized in a very smooth way, for example, by treatment with 0.05-0.5% Triton X-100 during 1-15 minutes after fixation with 0.5-1% glutaraldehyde in PBS-buffer (10 mM phosphate buffer; 150 mM NaCl, pH 7.4) or with 0.1% saponin in PHEM-buffer (1 mM MgSO₄; 10 mM EGTA; 60 mM PIPES; 25 mM HEPES; pH 6.9) during 1-15 minutes after fixation with 0.5-1% glutaraldehyde.

Under similar conditions, some aggregates of the present invention even penetrate into the nuclei of eukaryotic cells such as, PTK-2 cells.

Further it should be noted that in gel-filtration experiments, with for example Ultro gel ACA 44® (LKB), the specific binding agents being bound with one or more ultra small metal colloidal particle behave in a similar way as uncoupled specific binding agents, whereas aggregates based on larger colloidal metal particles behave as high-molecular weights.

As substances which can be detected according to the invention there may be mentioned any substance for which a specific binding agent exists. For example such substances comprise but are not limited to cell surface and tissue antigens, receptors, biological substances excreted by or derived from living organisms, particularly biological substances occurring in biological fluids such as saliva, lymph, blood and its derived fractions such as, plasma and serum, urine, cerebrospinal fluid, amnion fluid, and the like. Substances which can be detected include, proteins, polypeptides, peptides, like enzymes, hormones, structural proteins, receptors, nucleic acids, vitamins, polysaccharides, toxins, alkaloids, glycoproteins, haptens, metabolites, pharmacological agents, pesticides, pollutants, steroids, and any other molecule for which a specific binding counterpart exists in biological systems or can be synthesized.

The specific binding agents which can be used according to the invention can be of various nature but will in many instances be antibodies to specified antigens or haptens. As antibodies there may

be mentioned, immunoglobulins such as, IgM, IgG, IgA, IgD, and IgE, IgG being preferred, and their fragments, e.g., Fc, Fab and F(ab)₂. Antibodies may be polyclonal or monoclonal. As an example of specific binding substances other than antibodies there can be mentioned Staphylococcus Aureus protein A which specifically binds immunoglobulins of various animal species, protein G, streptavidine and avidine, other bindable substances may be phages, which are optionally chemically or genetically adapted to bind molecular or cellular materials, lectins, which specifically bind glycoproteins, DNA or RNA probes for gene identification, or fragments thereof and drugs which have a sufficient specificity and affinity for receptors. In general any other molecular interaction of sufficient specificity and affinity can be employed.

Representative protein analytes include the classes of protamines, mucoproteins, glycoproteins, globulins, albumins, scleroproteins, phosphoproteins, histones, lipoproteins, chromoproteins, and nucleoproteins. Examples of specific proteins are prealbumin, α_1 -lipoprotein, human serum albumin, α_1 -acid glycoprotein, α_1 -antitrypsin, α_1 -glycoprotein, transcortin, thyroxine binding globulin, haptoglobin, hemoglobin, myoglobin, ceruloplasmin, α_2 -lipoprotein, α_2 -macroglobulin, β -lipoprotein, erythropoietin, transferrin, hemopexin, fibrinogen, the immunoglobulins such as IgG, IgE, IgM, IgA, IgD, and IgE, IgG being preferred and their fragments, e.g., Fc, Fab and F(ab)₂ complement factors, prolactin, blood clotting factors such as fibrinogen, thrombin and so forth, insulin, melanotropin, somatotropin, thyrotropin, follicle stimulating hormone, luteinizing hormone, gonadotropin, thyroid stimulating hormone, placental lactogen, intrinsic factor, transcobalamin, serum enzymes such as alkaline phosphatase, lactic dehydrogenase, amylase, lipase, phosphatases, cholinesterase, glutamic oxaloacetic transaminase, glutamic pyruvic transaminase, and uropepsin, endorphins, enkephalins, protamine, tissue antigens, bacterial antigens, and viral antigens such as hepatitis associated antigens (e.g., HBsAg, HBeAg and HBeAg).

Representative hapten analytes include the general classes of drugs, metabolites, hormones, pesticides, pollutants, vitamins, and the like organic compounds. Haptenic hormones include thyroxine and triiodothyronine. Vitamins include vitamins A, B, e.g. B₁₂, C, D, E and K, folic acid and thiamine. Drugs include antibiotics such as aminoglycosides, e.g., gentamicin, tobramycin, amikacin, sisomicin, kanamycin, and netilmicin, penicillin, tetracycline, terramycin, chloromycetin, and actinomycetin; nucleosides and nucleotides such as adenosine diphosphate (ADP) adenosine triphosphate (ATP), flavin mononucleotide (FMN), nicotinamide adenine

dinucleotide (NAD) and its phosphate derivative (NADP), thymidine, guanosine and adenosine; prostaglandins; steroids such as the oestrogens, e.g., oestriol and oestradiol, steroids; and others such as phenobarbital, phenytoin, pirimidine, ethosuximide, carbamazepine, valproate, theophylline, caffeine, propranolol, procainamide, quinidine, amitriptyline, cortisol, desipramine, disopyramide, doxepin, doxorubicin, nortriptyline, methotrexate, imipramine, lidocaine, N-acetyl-procainamide, amphetamines, catecholamines, and antihistamines. Further cardiac glycosides, and derivatives of benzodiazepine, benzimidazole, piperidine, piperazine, imidazole, triazole, pyridazine, 1,2,4-triazinedione or 2,3,5,6-tetrahydro-imidazo[2,1-b]thiazoles, or amides, hydratropic acid derivatives or trialkylamines.

Benzimidazole haptens comprise thiabendazole, fuberidazole, cicloabendazole, oxiabendazole, parbendazole, cambendazole, mebendazole, fenbendazole, flubendazole, albendazole, oxfendazole, nocodazole and astemizole.

Piperidine haptens comprise diphenoxylate, phenoperidine, haloperidol, haloperidol decanoate, bromperidol decanoate, bromperidol, moperone, triflupridol, pipamperone, piritramide, fentanyl, buprenorphine, droperidol, benzitramide, benzetimide, domperidone, sufentanil, carfentanil, alfentanil, dexetomidine, milnaperone, difenoxin, fluspirilene, penfluridol, pimozide, lorcanide, loperamide, astemizole, ketanserine, levocabastine, cisapride, alantaserin, ritanserine, 3-[2-[4-(4-fluorobenzoyl)-1-piperidinyl]-ethyl]-2,7-dimethyl-4H-pyrido[1,2-a]pyrimidin-4-one, 3-[2-[4-[bis(4-fluoro-phenyl)-methylene]-1-piperidinyl]-ethyl]-2-methyl-4H-pyrido[1,2-a]pyrimidin-4-one and 3-[2-[4-[(3-furanylmethyl)-3H-imidazo[4,5-b]pyridin-2-yl]-amino]-1-piperidinyl]-ethyl]-2-methyl-4H-pyrido[1,2-a]pyrimidin-4-one.

Piperazine haptens include azaperone, fluanisone, lidoflazine, flunarizine, mianserine, oxatomide, mifloflazine, clocinizine and cinnarizine.

Examples of imidazole haptens are metronidazole, ornidazole, ipronidazole, tinidazole, isoconazole, nimorazole, miconazole, burimamide, metiamide, metomidate, enilconazole or imazalil, etomidate, econazole, clotrimazole, carnidazole, cimetidine, doconazole, sulconazole, parconazole, orconazole, butoconazole, triadinole, tioconazole, valconazole, fluotrimazole, ketoconazole, oxiconazole, lombazole, bifonazole, oxmetidine, fenticonazole, tubulazole and (Z)-1-[2-chloro-2-(2,4-dichlorophenyl)-ethenyl]-1H-imidazole. Triazole haptens comprise virazole, azaconazole, etaconazole, propiconazole, penconazole, itraconazole and terconazole. Pyridazine haptens comprise for example, 3-chloro-6-[3,6-dihydro-4-(3-methylphenyl)-1-(2H)-pyridinyl]-pyridazine, 3-methoxy-6-[4-(3-

methyl-phenyl)-1-piperazinyl]pyridazine and the compounds of EP-A-156,433. 1,2,4-Triazinediones comprise for example, 2-chloro- α -(4-chlorophenyl)-4-(4,5-dihydro-3,5-dioxo-1,2,4-triazin-2(3H)-yl)-benzeneacetonitrile, 2,6-dichloro- α -(4-chlorophenyl)-4-(4,5-dihydro-3,5-dioxo-1,2,4-triazin-2(3H)-yl)benzeneacetonitrile and the compounds of EP-A-170,316.

Trialkylamines are, for example, diisopropylamine, propylamine. 2,3,5,6-Tetrahydro-imidazo[2,1-b]thiazoles comprise, for example, tetramisole or levamisole. Amides comprise for example, closantel, ambucetamide, isopropamide, uzepide metiodide, dextromoramide. A hydratropic acid hapten is, for example, suprofen.

The binding reaction will in almost all cases be allowed to proceed under mild conditions. The reaction mixture will in general be an aqueous medium with any desirable organic co-solvents being present in minor amounts. The temperature of the reaction will be maintained at a constant level in normal circumstances throughout the incubation period. Temperatures will generally be between 0 and 50°C, more usually between 20 and 40°C. Preferably the reaction will proceed at room temperature. The pH of the reaction mixture will vary between 5 and 10, more usually between 6 and 9. The concentration of various reagents will depend on the level of analyte expected in the test medium, with such level usually being between 10^{-3} and 10^{-12} M. As in the case of the previously described reaction parameters, election is primarily based on empirically derived optimization balanced against the preferences and needs of the technician who will ultimately perform assays on a routine basis. None of the parameters therefore is of a critical nature to the present invention, rather they are all within the ordinary ranges used in the art.

The aggregates of the present invention can be employed in virtually all circumstances for which immunological techniques are conceived at present, especially those techniques which in general apply colloidal metal particles coupled to well defined organic substances. However, due to the penetrative capacities of the present aggregates the use will not be limited to those instances where at present metal labelling is known to be useful. In principle it can be applied to the qualitative and/or quantitative determination of any substance for which specific binding agents exist.

The aggregates of the present invention are particularly useful in those areas using specific detection techniques. For example in light or electron microscopic applications or blotting techniques, optionally in combination with chemical enhancement techniques e.g. silver enhancement, electronic enhancement techniques, electron energy loss detection techniques and/or X-ray micro

analysis.

Appropriate silver enhancement techniques are described in for example, the EP-A 165,634, in EP-A-239947 and in Histochemistry 85, 209-214 (1986). Appropriate electronic enhancement techniques are for example described in U.S. Pat. No. 4,752,567.

In light microscopical applications the present aggregates are especially useful for the immunological detection of antigens in combination with chemical enhancement, particularly silver enhancement, both in cell and tissue sections and in whole mount (intact cell) specimens. The thus achieved sensitivities are very high due to the fact that a large number of antigens are reachable for the penetrating aggregate and a higher labelling density is obtained as a result of the reduced steric hinderance. For example, the results for labelling tubulin are comparative or better than those which were achieved on basis of the same specific antibody labelled with an immunofluorescence marker. Besides, the signal generated is permanent and not prone to rapid degradation and no special microscopic equipment is needed. The resulting labelling after silver enhancement is particulate and can be quantified.

In electron microscopical applications the present aggregates are especially suited for high-resolution studies on distinct macromolecules or fragments thereof. The fact that the aggregates according to the present invention are capable of penetrating in standard biological specimens without any special pre-treatment makes them particularly useful in electron microscopical applications, e.g., to study the substructure of living material such as cells. In combination with chemical enhancement, they are suitable for immunolocalisation experiments, such as, preembedding immunolabelling of cells or tissue slices; post-sectioning labelling on plastic embedded material or cryosections; and labelling on whole mount cells. Preembedding labelling after mild permeabilization provides for efficient labelling with high sensitivity (penetration and efficiency), combined with optimized ultrastructural preservation. The resulting labelling after silver enhancement is particulate and can be quantified.

In blotting applications the present aggregates are especially suited for the identification of proteins in electrophoretically separated protein patterns. Compared to existing aggregates the detection is more sensitive and the resulting staining pattern is more delineated.

Due to their general character the aggregates of the invention, if desired, in combination with an enhancement technique the aggregates may further find utility in a variety of diagnostic tests such as, for example, the following: the detection and characterisation of subpopulations of T-lym-

phocytes; pregnancy tests based on the presence of certain hormones (chorionic gonadotropin) in the urine, diagnostic tests for various infections diseases of i.a. fungal, bacterial and in particular viral origin such as, for example, hepatitis B, autoimmune-diseases (e.g. AIDS), gonorrhoea, rubella, poliomyelitis, and the like, diagnostics for metabolic, endocrinological and various endogenous diseases, including diagnostics for the detection of congenital malfunctions of embryos based on the presence of specific proteins in the amnion fluid.

The invention further comprises reagent systems which comprise all of the essential chemical elements required to conduct a desired assay method encompassed by the present invention. The reagent system can be presented in a commercially packaged form, as a composition or admixture where the compatibility of the reagents will allow it, in a test device configuration, or as a test kit, i.e. a packaged combination of one or more containers holding the appropriate reagents. Included in the reagent system may be the reagents appropriate for the binding reaction system desired. Such binding reaction reagents can include, in addition to the labeled reagent, a binding counterpart to the analyte, and so forth. Of course, the reagent system can include other reagents as are known in the art and which may be desirable from a commercial and user stand-point, such as buffers, diluents, standards, and so forth.

Example 1

Preparation of colloidal gold dispersion containing ultra-small gold particles.

In a 1 litre beaker 200 ml distilled water was rapidly stirred at 300 r.p.m. and at a temperature of 4°C. 1.2 ml of a solution of ethanol, saturated at 4°C with white phosphorous was added over 15 seconds below the surface of the water, followed immediately by the addition of 200 ml of 0.005-0.01 % aqueous tetrachlorauric acid. Completion of the reduction was indicated by the appearance of a brown color. The excess of the reduction agent was subsequently removed by vigorously stirring the sol under reduced pressure with a water jet pump during 5 minutes after the sol was adjusted to pH 11.5 with sodium hydroxide at 4°C.

Example 2

Preparation of colloidal gold-labelled anti-rabbit immunoglobulin (Ig) antibodies.

Affinity-purified goat-anti-rabbit Ig antibodies were dialysed against 5 mM NaHCO₃ pH 9.8 during 24 hours at 4°C. The acidity of the gold sol

was brought to pH 9.0 with 0.2 M K_2CO_3 solution. 100 ml of this gold sol was stirred in a beaker at room temperature. 5 mg of the dialyzed antibodies were added to obtain a final concentration of 50 μ g/ml. 1 Minute later, 1 g bovine serum albumin (BSA) was added predissolved in 10 ml 1 mM sodium hydroxide. The thus obtained aggregate was dialyzed by vacuum dialysis against TBS-buffer (20 mM Tris; 150 mM sodium chloride, 20 mM NaN_3 ; pH 8.2).

Example 3

Penetration experiment (intact cells)

PTK-2 cells were grown on coverslips. Isolated coverslips were washed with PBS-buffer (10 mM phosphate buffer; 150 mM sodium chloride; pH 7.4) and fixed and/or permeabilized according to either one of the following procedures:

Method A:

The cells were fixed and permeabilized with 0.5% glutaraldehyde in PBS-buffer and 0.2% Triton X-100 during 10 minutes. The cells were further permeabilized during 15 minutes with 0.5% Triton X-100 in PBS-buffer.

Method B:

The cells were fixed with 0.5% glutaraldehyde in PBS during 10 minutes and permeabilized during 15 minutes with 0.5% Triton X-100 in PBS-buffer.

Method C:

The cells were washed with PHEM-buffer (1 mM $MgSO_4 \cdot 7 H_2O$; 10mM EGTA; 60 mM PIPES; 25 mM HEPES; pH 6.9) and then fixed with a 0.5% glutaraldehyde solution in PHEM-buffer during 10 minutes. After washing with a PHEM-buffer, the cells were permeabilized with a 0.1% saponin solution in PHEM-buffer during 15 minutes and then washed again with PHEM-buffer.

Method D:

The cells were fixed with 0.5% glutaraldehyde in PBS-buffer. After the fixation and permeabilization, the specimens were treated with 0.1 M $NaBH_4$ in buffer (PBS or PHEM) during 15 minutes. The whole was washed successively with PBS- or PHEM-buffer (3 x 5 minutes) and with a TBG-buffer (20 mM Tris; 150 mM sodium chloride; 20 mM NaN_3 ; 0.1% (w/v) BSA; 0.1% (w/v) CWF-gelatin (Sigma G 7765) and saponin 0.1% if method C is used; pH 8.2). The

mixture was preincubated with 5% normal goat serum in TBG-buffer (+ saponin) during 20 minutes and incubated overnight with rabbit-anti-tubulin 1 mg/ml in TBG (+ saponin), supplemented with 1% normal goat serum. After washing three times, each 15 minutes, with TBG-buffer (+ saponin), the cells were incubated during 4 hours with secondary goat-anti-rabbit (IgG) antibody/colloidal gold aggregates (the antibody concentration is ± 1 mg/ml in TBG-buffer, 1% (w/v) normal goat serum). The whole was washed three times during 10 minutes with TBG buffer (+ saponin) and then three times during 10 minutes with PBS-buffer. After washing, the cells were postfixed with 2% (w/v) glutaraldehyde in PBS-buffer during 10 minutes. The whole was washed successively with PBS-buffer (3 x 10 minutes) and then with distilled water (3 x 1 minute). The contrast of the reaction complex was increased by silver amplification with Intense II® (Janssen Life Science products) for 15 minutes. The coverslips are dehydrated and embedded in Pertex® (Histo-Lab).

Results:

Goat-anti-rabbit (IgG) antibody/ultra small colloidal gold (± 1 nm) aggregates penetrated into cell specimens prepared according to the procedures A, B, C and D whereas goat-anti-rabbit (IgG) antibody/colloidal gold (± 5 nm) aggregates only penetrate into specimens prepared according to procedure A.

Example 4

Penetration of ultra small gold probes into semithin cryosections.

Strenght of labeling compared to section thickness.

CHEMICALS AND REAGENTS

Goat anti-rabbit IgG polyklonal antibodies were conjugated to ultra small gold particles as previously described. Normal goat serum, GAR-G5nm and Inten SEII were purchased from Janssen Biotech, Olen/Belgium. PIPES, HEPES, bovine serum albumine and cold-waterfish gelatin were obtained from Sigma, St. Louis MO. Glutaraldehyde from BDH, Poole/United Kingdom. All other chemicals were purchased from Merck, Darmstadt/FRG and were of PA grade. All culture fluids were obtained from Gibco Paisley/United Kingdom

CELL CULTURES

Chinese hamster ovary cells (CHO cells) were cultured in Eagle's minimal essential medium supplemented with 10% fetal bovine serum.

SPECIMEN PREPARATION

After release from nocodazole treatment (ref. De Brabander et al., Microtubules and microtubule inhibitors 1980 page 225-268; Cell Motility 1981 page 469-483), mitotic CHO cells in metaphases and telophases were fixed in 0.5% glutaraldehyde in 60 mM PIPES, 25 mM HEPES; 10 mM EGTA, 1 mM magnesium sulfate (PHEM buffer), pH 6.9 for 10 minutes.

Cells were embedded in 10% liquified gelatin at 37°C. Next the cells were pelleted by centrifugation at 325 g for 3 minutes (Eppendorf centrifuge).

After solidification at 4°C, the gelatin was fixed with 0.5% glutaraldehyde in PHEM buffer for 10 minutes. For cryoprotection, cell pellets were infused with 0.6-1.5-2.3 M sucrose in PBS.

After cryoprotection cell blocks were cut of 1-2 mm³.

CRYO-SECTIONING

Serial semithin cryosections (thickness 0.2 and 0.4 µm) were cut on a Reichert FC4D cryoultramicrotome. The sections were attached to coverslips and stored overnight in PBS at 4°C.

IMMUNOLABELING

Sections were treated with sodium borohydride (1mg/ml) in PBS for 15 minutes to reduce residual aldehyde groups. Next the sections were rinsed 3 x 5 minutes in PBS and 2 x 5 minutes PBS supplemented with 0.8 BSA, 0.1% cold water fish gelatin, 2 mM azide, pH 7.4 (PBG - buffer). Aspecific staining was blocked by incubating by the sections in 5% normal goat serum in PBG buffer for 20 minutes.

Next the sections were incubated with a rabbit polyklonal antibody against tubulin's (diluted to 1 µg/ml in PBG buffer supplemented with 1% normal goat serum) for 60 minutes. Detection was done either with a goat antirabbit polyklonal antibody conjugated to 5 nm gold or to the ultra small gold particles. The surplus of gold probe was washed away with PBG-buffer 3 x 15 minutes.

After rinsing the section in PBS, they were fixed in 2% glutaraldehyde in PBS for 20 minutes. Silver enhancement was used for the light microscopical visualisation of the gold signal. Specificity and method controls for the immunocytochemical staining were as follows : omission of the antitubulin antibody and/or the gold conjugated goat anti-rabbit IgG antibody. The sections were rinsed in dis-

tilled water and dehydrated in a graded series of ethanol.

After mounting, the preparations could be evaluated.

RESULTS

Sections incubated with the 5 nm gold conjugate and the ultra small gold conjugate both showed a staining of the microtubules and the bulk of non polymensed tubulin in the cytoplasm of CHO cells. Background staining was in both cases negligible. However, when the ultra small gold probe was used the specific signal in a 0.4 µm section was for more stronger than in a serial 0.2 µm section. This observation was in contrast with the one where the 5 nm gold conjugate was used. There the signal intensity remained the same.

Example 5

Penetration of ultra small gold probes into semithin cryosections. Transverse sectioning of immunogold labeled semithin cryosections.

Semithin cryosections (thickness 0.4 µm) of CHO cells embedded in gelatin, were attached to the bottom of plastic Petri dishes. They were labeled with a 5 nm gold probe or with the ultra small gold probe, as described in Example 4. The 5 nm gold labeled cryosections were further processed for electron microscopical evaluation without silver enhancement. Silver enhancement times for the sections labeled with the ultra small gold probe lasted up to 4 minutes.

PLASTIC EMBEDDING AND SECTIONING

After being immuno stained the semithin cryosections were dehydrated in a graded series of ethanol and embedded in Epon in the Petri-dishes. After polymerization at 50°C for 24 hours, the plastic dishes were removed and the sections were reembedded in flat molds in Epon for transverse sectioning.

Ultrathin plastic sections were cut on a Reichert ultracut E and picked up on grids with a carbon coated parlodion film. They were stained with uranyl acetate and lead citrate before examination in a Philips EM 410 electron microscope.

RESULTS

When treated properly, reembedded semithin cryosections showed a well preserved morphology although cracks and clefts could be seen.

Plastic cross sections through labeled and reembedded semithin cryosections allowed determina-

tion of the gold or goldsilver particles on or within the sections. After silver enhancement, the ultra small gold particles could be made clearly visible. A silver enhancement time of 2 minutes showed goldsilver particles approximating 5 nm.

Transverse sectioning showed that immunolabeling with 5 nm gold probes was confined to the surface of semithin cryosections if there was no visible damage. If there were cracks or clefts in the section, only on those sites the 5 nm particles could be seen inside the section. Semithin cryosections with a severely disrupted ultrastructure showed overall penetration of the 5 nm goldprobe.

Well preserved cryosections showed labeling with the ultra small goldprobe not only on the surface of the section, but also throughout the whole inside. Even cells with an intact outer cellmembrane showed an intense labeling. Damaged semithin cryosections were also labeled throughout.

Claims

1. An aggregate for determining bindable substances comprising a colloidal metal particle and a specific binding agent for said bindable substance characterized in that the mean diameter of said colloidal particle is ranging between 0.8 and 2 nm.
2. An aggregate for determining bindable substances comprising a colloidal metal particle and a specific binding agent for said bindable substance characterized in that the size of the metal particle is selected so that said aggregate is capable of penetrating into standard biological specimens without any special pre-treatment or which have been permeabilized by non-extensive treatment with detergents in such way that the biological structural coherence is not strongly destroyed.
3. An aggregate according to claim 2 wherein the standard biological specimens are permeabilized by treatment with 0.05-0.5% Triton®X-100 during 1-15 minutes after fixation with 0.5-1% glutaraldehyde in PBS-buffer (10 mM phosphate buffer; 150 mM NaCl, pH 7.4) or with 0.1% saponin in PHEM-buffer (1 mM MgSO₄; 10 mM EGTA; 60 mM PIPES; 25 mM HEPES; pH 6.9) during 1-15 minutes after fixation with 0.5-1% glutaraldehyde.
4. An aggregate according to claim 2 or 3 wherein the standard biological specimens are not specially pre-treated.
5. An aggregate according to any of claims 1 to 4 wherein the colloidal metal particles are col-

loidal gold, silver, platinum, palladium, thallium, copper or nickel particles and their radioactive isotopes.

- 5 6. An aggregate according to any of claims 1 to 4 wherein the mean ratio of the colloidal metal particle to the binding agent is at least one.
7. An aggregate according to any of claims 1 or 4 where in the specific binding agent is an antibody, avidin, streptavidin, protein A or protein G.
8. An aggregate according to claim 7 wherein the specific binding agent is an immunoglobulin or a fragment thereof.
9. A method for qualitatively or quantitatively determining bindable substances characterized by using an aggregate as claimed in any of claims 1 to 8.
10. A method according to claim 9 wherein the bindable substances are part of substructures in living material.
11. A process for preparing aggregates as claimed in any of claims 1 to 8, characterized by attaching said colloidal metal particles to specific binding agents.
12. An aggregate for determining bindable substances according to any of claims 1 to 8, comprising a colloidal metal particle and a specific binding agent for said bindable substance whereby the colloidal metal particle is obtainable by a homogeneous reduction of an appropriate salt, acid or complex of the required metal with a strong reductant at a temperature ranging between 0 and 10 °C.
13. An aggregate according to claim 12 wherein the homogeneous reduction comprises the steps of
 - i) preparing an aqueous solution of an appropriate salt, acid or complex of the required metal;
 - ii) preparing an aqueous suspension of phosphorous by mixing water with an appropriate amount of a saturated solution of phosphorous in a lower alkanol or an ether;
 - iii) quickly adding one of the solutions to the other under vigorous stirring to form a colloidal sol; and, if desired,
 - iv) removing the excess amount of phosphorous and/or the lower alkanol or ether from the sol at a temperature ranging between 0 and 10 °C.

14. An aggregate according to claim 13, wherein in step i) the salt, acid or complex is tetrachloroauric acid; in step ii) the lower alkanol is ethanol; step iii) is conducted at a temperature between 0 and 4 °C; in step iv) the removing of phosphorous and/or ethanol is done under reduced pressure after addition of a sufficient amount of base to convert the remaining phosphorous to phosphine.

15. A test kit for quantitatively or qualitatively determining bindable substances characterized in that it contains an aggregate as claimed in any of claims 1 to 8.

Patentansprüche

1. Aggregat zum Bestimmen bindungsfähiger Substanzen, welches ein kolloidales Metallteilchen und ein spezifisches Bindungsmittel für die genannte bindungsfähige Substanz umfaßt, dadurch gekennzeichnet, daß der mittlere Durchmesser des kolloidalen Teilchens in einen Bereich von 0,8 bis 2 nm fällt.

2. Aggregat zum Bestimmen bindungsfähiger Substanzen, welches ein kolloidales Metallteilchen und ein spezifisches Bindungsmittel für die genannte bindungsfähige Substanz umfaßt, dadurch gekennzeichnet, daß die Größe des Metallteilchens so ausgewählt ist, daß das Aggregat dadurch befähigt wird, in biologische Standardproben einzudringen, die entweder keiner besonderen Vorbehandlung unterworfen worden sind oder welche durch eine nicht-umfassende Behandlung mit Detergentien derart permeabilisiert worden sind, daß dadurch der biologische strukturelle Zusammenhalt nicht stark zerstört worden ist.

3. Aggregat nach Anspruch 2, wobei die biologischen Standardproben entweder durch eine Behandlung mit 0,05- bis 0,5%igem Triton® X-100 während 1 bis 15 min nach einem Fixieren mit 0,5- bis 1%igem Glutaraldehyd in PBS-Puffer (10 mM Phosphatpuffer; 150 mM NaCl, pH 7,4) oder durch eine Behandlung mit 0,1%igem Saponin in PHEM-Puffer (1 mM MgSO₄; 10 mM EGTA; 60 mM PIPES; 25 mM HEPES; pH 6,9) während 1 bis 15 min nach einem Fixieren mit 0,5- bis 1%igem Glutaraldehyd permeabilisiert worden sind.

4. Aggregat nach Anspruch 2 oder 3, wobei die biologischen Standardproben keiner speziellen Vorbehandlung unterworfen worden sind.

5. Aggregat nach einem der Ansprüche 1 bis 4,

wobei die kolloidalen Metallteilchen kolloidale Gold-, Silber-, Platin-, Palladium-, Thallium-, Kupfer- oder Nickelteilchen und deren radioaktive Isotope sind.

6. Aggregat nach einem der Ansprüche 1 bis 4, wobei das mittlere Verhältnis zwischen dem kolloidalen Metallteilchen und dem Bindungsmittel wenigstens 1 beträgt.

7. Aggregat nach einem der Ansprüche 1 bis 4, wobei des spezifische Bindungsmittel ein Antikörper, Avidin, Streptavidin, Protein A oder Protein G ist.

8. Aggregat nach Anspruch 7, wobei des spezifische Bindungsmittel ein Immunglobulin oder ein Fragment desselben ist.

9. Verfahren zum qualitativen oder quantitativen Bestimmen bindungsfähiger Substanzen, gekennzeichnet durch die Verwendung eines in einem der Ansprüche 1 bis 8 beanspruchten Aggregates.

10. Verfahren nach Anspruch 9, wobei die bindungsfähigen Substanzen ein Teil der Substrukturen in lebendem Material sind.

11. Verfahren zur Herstellung der in einem der Ansprüche 1 bis 8 beanspruchten Aggregate, dadurch gekennzeichnet, daß die kolloidalen Metallteilchen an spezifische Bindungsmittel gebunden werden.

12. Aggregat zum Bestimmen bindungsfähiger Substanzen, nach einem der Ansprüche 1 bis 8, welches Aggregat ein kolloidales Metallteilchen und ein spezifisches Bindungsmittel für die genannte bindungsfähige Substanz umfaßt, und wobei des kolloidale Metallteilchen durch eine homogene Reduktion eines entsprechenden Salzes, einer entsprechenden Säure oder eines entsprechenden Komplexes des erforderlichen Metalles mit einem starken Reduktionsmittel bei einer in einen Bereich von 0 bis 10 °C fallenden Temperatur erhältlich ist.

13. Aggregat nach Anspruch 12, wobei die homogene Reduktion die folgenden Stufen umfaßt:

- i) Herstellen einer wäßrigen Lösung eines entsprechenden Salzes, einer entsprechenden Säure oder eines entsprechenden Komplexes des erforderlichen Metalles;
- ii) Herstellen einer wäßrigen Phosphorsuspension durch Vermischen von Wasser mit einer entsprechenden Menge einer gesättigten Lösung von Phosphor in einem niede-

- ren Alkanol oder einem Ether;
 iii) rasches Zugeben einer der Lösungen zu der anderen unter kräftigem Rühren zur Bildung eines kolloidalen Sols; und, gewünschtenfalls,
 iv) Entfernen der überschüssigen Menge von Phosphor und/oder dem niederen Alkanol oder Ether von Sol bei einer Temperatur im Bereich von 0 bis 10° C.
14. Aggregat nach Anspruch 13, wobei in Stufe i) das Salz, die Säure oder der Komplex Tetrachlorgold(III)-säure ist; in Stufe ii) das niedere Alkanol Ethanol ist; die Stufe iii) bei einer Temperatur von 0 bis 4° C durchgeführt wird; in Stufe iv) des Entfernen von Phosphor und/oder Ethanol unter reduziertem Druck nach Zugabe einer für die Umwandlung des restlichen Phosphors zu Phosphin ausreichenden Menge einer Base vorgenommen wird.
15. Testkit zum quantitativen oder qualitativen Bestimmen bindungsfähiger Substanzen, dadurch gekennzeichnet, daß er ein in einem der Ansprüche 1 bis 8 beanspruchtes Aggregat enthält.

Revendications

1. Agrégat pour la détermination de substances liables comprenant une particule métallique colloïdale et un agent de liaison spécifique pour ladite substance liable caractérisé en ce que le diamètre moyen de ladite particule colloïdale s'étend entre 0,8 et 2 nm.
2. Agrégat pour la détermination de substances liables comprenant une particule métallique colloïdale et un agent de liaison spécifique pour ladite substance liable caractérisé en ce que la taille de la particule métallique est choisie de façon que ledit agrégat soit apte à pénétrer à l'intérieur de spécimens biologiques standards sans pré-traitement spécial quelconque ou qui a été perméabilisé au moyen d'un traitement non-extensive au moyen de détergents de telle sorte que la cohérence de la structure biologique ne soit pas fortement détruite.
3. Agrégat selon la revendication 2, dans lequel les spécimens biologiques standards sont perméabilisés par traitement avec le Triton® X-100 à 0,05-0,5% pendant 1 à 15 minutes après fixation avec la glutaraldéhyde à 0,5-1% dans le tampon PBS (tampon phosphate 10mM; NaCl 150 mM, pH 7,4) ou avec une saponine à 0,1% dans le tampon PHEM (MgSO₄ 1mM; EGTA 10 mM, PIPES 60 mM; HEPES 25 mM; pH 6,9) pendant 1 à 15 minutes après fixation avec la glutaraldéhyde à 0,5 à 1%.
4. Agrégat selon la revendication 2 ou 3, dans lequel les spécimens biologiques standard ne sont pas pré-traités en particulier.
5. Agrégat selon l'une quelconque des revendications 1 à 4, dans lequel les particules métalliques colloïdales sont des particules d'or, d'argent, de platine, de palladium, de thallium, de cuivre ou de nickel colloïdales et leurs isotopes radioactifs.
6. Agrégat selon l'une quelconque des revendications 1 à 4, dans lequel le rapport moyen de la particule métallique colloïdale à l'agent de liaison est d'au moins un.
7. Agrégat selon l'une quelconque des revendications 1 à 4, dans lequel l'agent de liaison spécifique est un anti-corps, une avidine, une streptavidine, une protéine A ou une protéine G.
8. Agrégat selon la revendication 7, dans lequel l'agent de liaison spécifique est une immunoglobuline ou un fragment de celle-ci.
9. Procédé de détermination qualitative ou quantitative de substances liables caractérisé par l'utilisation d'un agrégat selon l'une quelconque des revendications 1 à 8.
10. Procédé selon la revendication 9, dans lequel les substances liables sont une partie de sous-structures dans une substance vivante.
11. Procédé de préparation d'agrégat selon l'une quelconque des revendications 1 à 8 caractérisé par la liaison desdites particules métalliques colloïdales aux agents de liaison spécifiques.
12. Agrégat pour la détermination de substances liables selon l'une quelconque des revendications 1 à 8 comprenant une particule métallique colloïdale et un agent de liaison spécifique pour ladite substance liable d'où l'on peut obtenir la particule métallique colloïdale au moyen d'une réduction homogène d'un sel, d'un acide ou d'un complexe approprié du métal nécessaire avec un réducteur fort à une température s'étendant entre 0 et 10° C.
13. Agrégat selon la revendication 12, dans lequel la réduction homogène comprend les étapes

de

i) préparation d'une solution aqueuse d'un sel, d'un acide ou d'un complexe approprié du métal nécessaire;

ii) préparation d'une suspension aqueuse de phosphore en mélangeant de l'eau avec une quantité appropriée d'une solution saturée de phosphore dans un alcanol inférieur ou un éther; 5

iii) addition rapide d'une des solutions à l'autre sous agitation vigoureuse pour former un sol colloïdal; et, si souhaité, 10

iv) élimination de la quantité en excès du phosphore et/ou de l'alcanol inférieur ou un éther du sol à une température s'étendant entre 0 et 10° C. 15

14. Agrégat selon la revendication 13, dans lequel dans l'étape i) le sel, l'acide ou la complexe est l'acide tétrachloroaurique; dans l'étape ii) l'alcanol inférieur est l'éthanol; l'étape iii) est conduite à une température entre 0 et 4° C; dans l'étape iv) l'élimination du phosphore et/ou de l'éthanol est effectuée sous pression réduite après addition d'une quantité suffisante de base pour convertir le phosphore restant en phosphine. 20 25

15. Trousse de test pour la détermination quantitative ou qualitative de substances liables, caractérisée en ce qu'elle contient un agrégat selon l'une quelconque des revendications 1 à 8. 30

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